RESEARCH

Open Access

Sustained delivery of chABC improves functional recovery after a spine injury



Atousa Sharifi¹, Ali Zandieh¹, Zahra Behroozi^{2,6}, Michael R. Hamblin³, Sara Mayahi⁴, Mahmoud Yousefifard⁵ and Fatemeh Ramezani^{5*}

Abstract

Introduction: Chondroitinase ABC (chABC) is an enzyme could improve regeneration and thereby improving functional recovery of spinal cord injury (SCI) in rodent models. Degradation of the active enzyme and diffusion away from the lesion are the causes of using hydrogels as a scaffold to deliver the chABC into the lesion site. In this meta-analysis, we investigated the effects of chABC embedded in a scaffold or hydrogel on the functional recovery after SCI.

Method: Databases were searched based on keywords related to chABC and spinal cord injury (SCI). Primary and secondary screening was performed to narrow down study objectives and inclusion criteria, and finally the data were included in the meta-analysis. The standard mean difference of the score of the functional recovery that measured by Basso, Beattie, Bresnahan (BBB) test after SCI was used to analyze the results of the reported studies. Subgroup analysis was performed based on SCI model, severity of SCI, transplantation type, and the follow-up time. Quality control of articles was also specified.

Results: The results showed that embedding chABC within the scaffold increased significantly the efficiency of functional recovery after SCI in animal models (SMD = 1.95; 95% CI 0.71-3.2; p = 0.002) in 9 studies. SCI model, severity of SCI, injury location, transplantation type, and the follow-up time did not affect the overall results and in all cases scaffold effect could not be ignored. However, due to the small number of studies, this result is not conclusive and more studies are needed.

Conclusion: The results could pave the way for the use of chABC embedded in the scaffold for the treatment of SCI and show that this method of administration is superior to chABC injection alone.

Keywords: chABC, Scaffold, Hydrogel, Spinal cord injury: animal model

Introduction

Spinal cord injury (SCI) is caused by damage to the spinal cord that causes is associated with temporary or permanent impairment in neurological function [1, 2]. SCI is divided into two types: traumatic and non-traumatic. Traumatic SCI occurs because of motor vehicle accidents, falls, sports injuries, violent assault, etc. While

Atousa Sarifi and Ali Zandie are co-first autors

*Correspondence: Ramezani.f@iums.ac.ir

non-traumatic SCI is produced by diseases such as tumors, infections, etc. [3, 4]. Traumatic SCI often lead to devastating loss of sensory and motor function, and despite therapeutic methods which have shown positive results in the research phase [5–10] so far there is no drug treatments that can restore function consistently in affected patients Although there is some spontaneous regenerative responses that occurs following SCI, this is often insufficient for significant improvement [11]. Several factors, including inappropriate immune responses, glial scar development, and lack of adequate neurotrophic support after SCI may be the cause of this



© The Author(s) 2022. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/. The Creative Commons Public Domain Dedication waiver (http://creativecommons.org/publicdomain/zero/1.0/) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

⁵ Physiology Research Center, Iran University of Medical Sciences, Tehran, Iran Full list of author information is available at the end of the article

lack of regenerative ability [12–15]. The glial scar and the cystic cavities that are formed over the long term tend to decrease remyelination and inhibit axonal regrowth [16]. As a result, the healing of the spinal cord is very poor compared to other types of injured tissue [17, 18].

Several experiments have shown that at the site of the lesion, chondroitin sulfate proteoglycans (CSPGs) are secreted in increasing quantities by the stimulated astrocytes and oligodendrocytes. These CSPGs subsequently inhibit the axonal regeneration that might occur after SCI [19–21]. The glial scar is also a physical barrier to axonal regeneration. CSPGs are inhibitory molecules that accumulate at the site of the lesion and prevent neurite extension, neuronal growth, and neuroplasticity both in vitro and in the injured CNS in-vivo [12, 22–25]. CSPGs is an extracellular matrix component that contains two common structural parts; one is the protein core (NG2 protein), and the attached long glycosaminoglycan (GAG) polysaccharide chains [26–29].

Chondroitinase ABC (chABC) is an enzyme that can break the sulfated GAG chains on the CSPGs which are a main element of the glial scar [30–33]. Degradation of the glial scar has been shown to stimulate axonal growth, leading to improved function in some models of SCI in rodents [17, 34–36]. chABC also degrades the CSPGs that form perineuronal nets, thereby enhancing axonal sprouting and plasticity [37–39]. However, the use of chABC in animal models has problems that have caused the effect of chABC to be moderately reported in a metaanalysis study based on the results of 34 preclinical studies [40].

One major issue is that the administration of chABC by injection, by using an intrathecal catheter, or pumping through a cannula cannot guarantee sufficient sustained concentrations of drug at the injury site [23, 30]. Moreover, because chABC is a protein it remains active for only 3-5 days at $37 \ ^{\circ}C \ [41-43]$.

One approach to prevent premature degradation of the enzyme and to increase its efficacy is to use a variety of scaffolds that can be loaded with chABC for sustained release [30, 43, 44]. These scaffolds are often hydrogels

Page 2 of 8

that possess low toxicity and are also biocompatible and biodegradable [45–47]. In various studies, chABC has been loaded into hydrogel scaffolds, and the effect on the treatment of SCI has been investigated in animal models. Some of these scaffolds have included PLGA nanoparticles [48], PLLA microspheres incorporated within a chitosan scaffold [49], other types of hydrogel such as methylcellulose hydrogel [30], agarose hydrogel-microtube scaffold system [41], cross-linked methylcellulose (XMC) hydrogel, collagen scaffold [43], or alginate electrospun scaffold [50].

In this meta-analysis, we compared the effects of chABC when loaded inside a scaffold with untreated animals, on functional improvements after SCI in animal models.

Material and methods

The guidelines of Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) were used for this study. The review protocol for this study has not been published.

Search strategy

Using keyword groups related to chABC and spinal cord injury, listed in Table 1, a systematic search with unlimited language limitation for published articles up to June 30, 2022, was performed in the databases of SCOPUS, EMBASE, MEDLINE, and Web of Science. A manual exploration was also accompanied to find further articles.

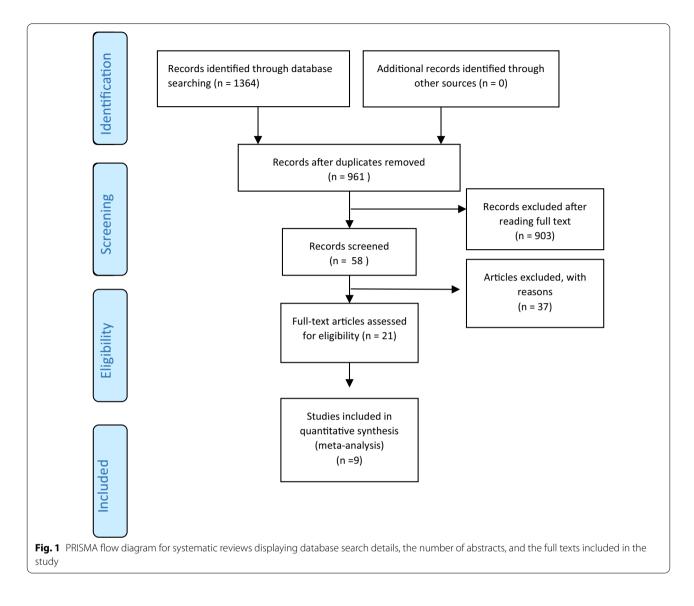
Study selection, eligibility, and rejection standards

Duplicate articles were removed and then the two reviewers reviewed the articles separately by reading the titles, abstracts, and complete texts (if the titles and abstracts did not cover necessary information). Disagreements over selection of studies were resolved through discussions between them.

Only studies were included for analysis that reported the effect of chABC embedded in any type of scaffold, using in-vivo experiments in animals with SCI, and the

Table 1 Search strategies used in PUBMED

PUBMED ("Chondroitin-Sulfate-ABC Endolyase" [MESH] or "Chondroitin Sulfate ABC Endolyase" [MESH] or "Chondroitinase ABC" [MESH] or "Chondroitin-Sulfate-ABC Endolyase" [tiab] or "Chondroitin Sulfate ABC Endolyase" [tiab] or "Chondroitinase ABC" [tiab] or "Chondroitin sulfate ABC endolyase" [tiab] or "Chondroitin sulfate ABC lyase" [tiab] or "Chondroitin sulfate ABC endolyase" [tiab] or "Chondroitin sulfate ABC endolyase" [tiab] or "Chondroitin sulfate ABC lyase" [tiab] or "Chondroitinase ABC" [tiab] or "Chondroitin sulfate ABC lyase" [tiab] or "Chondroitinase ABC" [tiab] or "Chondroitin sulfate ABC endolyase" [tiab] or "Chondroitin sulfate ABC lyase" [tiab] or "Chondroitin sulfate ABC endolyase" [tiab] or "Chondroitin sulfate ABC lyase" [tiab] or "Chondroitinase ABC" [tiab] or "Chondroitin sulfate ABC lyase" [tiab] or "Chondroitinase ABC" [tiab] or "Chondroitin sulfate ABC lyase" [tiab] or "Chondroitinase ABC" [tiab] or "Chondroitin sulfate ABC lyase" [tiab] or "Chondroitinase ABC" [tiab] or "Chondroitinse ABC" [tiab] or "Chondroitin sulfate ABC lyase" [tiab] or "Chondroitinse ABC" [tiab] or "Chondroitinse ABC" [tiab] or "Chondroitinse ABC" [tiab] or "Chondroitinse ABC" [tiab] or "Chondroitin sulfate ABC lyase" [tiab] or "Chondroitinse ABC" [tiab] or "Chondroitinse ABC" [tiab] or "Chondroitinse ABC" [tiab] or "Chondroitinse ABC" [tiab] OR "spinal cord lyase" [tiab] OR "spinal cord transection" [[tiab] OR "spinal cord transection" [tiab] OR "spinal cord transection" [tiab] OR "spinal cord transection" [tiab] OR "spinal cord "[tiab] OR "spinal cord" [tiab] OR "spinal cord" [tiab] OR "spinal cord "spinal cord" [tiab] OR "sp



result using the BBB test to measure functional recovery was compared with the untreated SCI group.

Exclusion criteria were: review articles, articles that did not use chABC embedded in scaffolding as intervention, studies that did not compare the treatment group with the control group.

Data extraction and quality assessment

Both assessors independently extracted the following information from the articles: animal features, species, gender, weight, age, sample size, studied organ, duration of exposure, study time, type of transplantation of chABC (implant or injection), chABC concentration, vehicle name and the score of BBB test which represent the functional recovery after SCI. Analysis was performed by calculation of standard mean differences (SMD) of the mean of BBB test and the reported standard deviation for each group. The risk of bias assessment of articles was achieved based on the study of Hassannejad et al. [51].

Statistical analysis

For data analysis STATA 14.0 statistical software was used. Data as mean and standard deviation were presented. The effect size was calculated with a 95% confidence interval (95% CI). The fixed-effect model was used, and if the heterogeneity was more than 50%, the random-effect model was used. Using Egger's precision-weighted linear regression method, the existence of publication bias was explored, and the results were available in funnel plots. Significant level in all analyses was considered p < 0.05.

Refs.	Gender; species; strain; weight or age	Model of injury; location of injury; severity	Numbers: sham; chABC alone; scaffold chABC	Interval time from injury to treatment	chABC concentration	Transplantation type (implant or injection)	Nanoscaffold or hydrogel	FU (week)
Azizi et al. [48]	Male; rat; Wistar; 275 ± 25 g	Contusion; T10; severe	5; NA; 6	1 week	%1 W/W	Injection	PLGA nanoparticles	8
Führmann et al. [53] Female; rat; SD; 300 g	Female; rat; SD; 300 g	Compression; T1-2; moderate	n ≥ 9; n ≥ 9; n ≥ 9	chABC immediately; cell injection 1 week later	5 µl	Injection	Methylcellulose (XMC) hydrogel	σ
Cholas et al. [52]	Female; rat; Lewis; 150–175	Hemiresection; T8-9; severe	6; NA; 8	0 day	25 µL of a 10-Units/ mL chABC	Implant	EDAC-cross-linked collagen scaffold	4
Pakulska et al. [30]	Female; rat; SD; 200–250 g	Compression; T1-2; moderate	11; 11; 11	Immediately after injury	10 µg	Injection	Methylcellulose hydrogel	œ
Xia et al. [54]	Female; rat; Wistar; 200–230 g	Hemitransection; T7–T9; moderate	6; 6; 6	During the proce- dure	NR	Implant	Poly(propylene car- bonate) microfibers scaffold	4
Zhang et al. [55]	Female; rat; SD; 200–250 g/2–3 mo	Segmental trans- versal injury; T10; severe	8; 8; 8	After surgery	0.25 U	Injection	PLGA delayed- release micro- spheres	10
Pan et al. [56]	Female; rat; Wistar; 250 g	Transection; T10; severe	8; 12; 12	Immediately after injury	10 U/mL	Implant	Poly(glycerol seba- cate)	12
Ni et al. [57]	Female; rat; Wistar; 200–230 g	Hemisection; T7–T9; severe	6; 6; 6	Immediately after injury	W/W 1%	Implant	Poly (propylene carbonate)-chitosan micron fibers scaf- fold	4

Table 2 Data of the studies included in the systematic review and meta-analysis

FAQ self assembled 6 weeks after injury peptide

Injection

0.1 µ/ml

4 weeks after injury

8; 8; 8

Compression; T9— T10, moderate

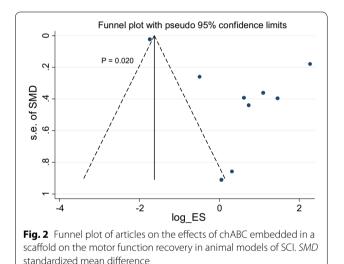
Female; rat; SD; 250–275 g

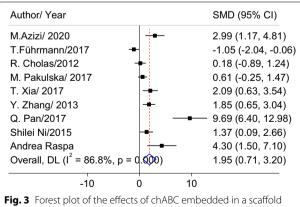
Raspa et al. [33]

Table 3 Bias items in all included studies

Author	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16
M. Azizi	Y	Y	Y	Y	Y	Y	Y	Y	Y	Y	Y	Y	Y	Y	Ν	N
T. Führmann	Y	Y	Y	Y	Y	Y	Y	Y	Y	Υ	Ν	Υ	Y	Υ	Y	Ν
R. Cholas	Y	Y	Y	Υ	Υ	Y	Y	Y	Y	Υ	Ν	Υ	Υ	Y	Ν	Ν
M. Pakulska	Y	Y	Y	Υ	Υ	Y	Y	Y	Y	Ν	Ν	Ν	Υ	Y	Ν	Ν
T. Xia	Y	Y	Y	Y	Y	Y	Y	Y	Y	Υ	Ν	Υ	Υ	Υ	Ν	Ν
Y. Zhang	Y	Y	Y	Y	Y	Y	Y	Y	Y	Ν	Υ	Υ	Υ	Ν	Y	Y
Q. Pang	Y	Y	Y	Υ	Υ	Y	Y	Y	Y	Y	Υ	Ν	Υ	Ν	Ν	Ν
Shilei Ni	Y	Y	Y	Υ	Y	Y	Y	Y	Y	Y	Ν	Υ	Υ	Y	Ν	Ν
Andrea Raspa	Y	Y	Y	Y	Y	Y	Y	Y	Y	Y	Ν	Ν	Y	Y	Ν	Ν

1. Species, 2. Strain, 3. Age/weight, 4. Genetic background, 5. Number of animals per group, 6. Definition of control, 7. Method of allocation to treatments, 8. Target tissue, 9. Using appropriate tests, 10. Blinding the assessor, 11. Randomization 12. Definition of the experimental unit (individual animal/animals in one cage), 13. Description of statistical analysis, 14. Animal facility, 15. Regulation and ethics, 16. Description of the reasons to exclude animals during the study (attrition)





on behavioral tests after SCI compared to non-treated animals. *CI* confidence interval, *SMD* standardized mean difference

Results

Included studies

1364 articles were initially acquired from a general search of the records. When removing identical articles, 961 articles were carefully chosen based on evaluation of the titles and abstract. After evaluating the full text, 9 articles finally were used for the meta-analysis. Figure 1 displays the flow chart of the search procedure and the choice of articles.

Data extraction

9 studies look at the effect of chABC embedded in a scaffold on animal locomotion after SCI. The data gained from these articles is prepared in Table 2. All 9 articles used a rat model. Two studies were performed on the T1-T2 location and the others were performed on T7– T10. All 9 articles used the BBB test to measure the locomotor activity of the animals. Therefore, a meta-analysis was performed to explore the effect of chABC embedded in a scaffold on the BBB test results.

Quality control

The calculation of the risk of bias indicated low risk of bias in all articles in the following categories: species, strain, genetic background, age/weight, and number of animals per group, description of control, method of group allocation, objective tissue, use of suitable tests, and description of statistical analysis.

2 articles displayed a great risk of bias regarding the blinding of the assessor, 6 articles displayed a high risk of bias regarding randomization, 2 articles displayed a high risk of bias in terms of the definition of the experimental unit, 2 articles showed a high risk of bias in terms of description of the animal facility, 7 articles showed a high risk of bias in term of regulations and ethics, and 8

Subgroup	Number of experiments	Heterogeneity (p value)	SMD (959	p value		
SCI model						
Contusion-compression	4	87.3% (< 0.0001)	1.4	- 0.52	3.31	0.15
Transection-hemisection	5	87% (<0.0001)	2.458	0.688	4.228	0.006
Severity of SCI						
Severe	5	87.8% (< 0.0001)	2.694	0.793	4.595	0.005
Moderate	4	85.7% (0.001)	1.146	- 0.567	2.858	0.19
Transplantation type						
Injection	5	85.7% (< 0.0001)	1.45	- 0.093	2.997	0.065
Implant	4	90.0% (< 0.0001)	2.801	0.399	5.204	0.022
Follow-up time						
< 8 weeks	6	67.2% (0.0091)	1.55	0.572	2.527	0.002
\geq 8 weeks	3	95.5% (< 0.0000)	3.11	- 0.89	7.1	0.128
Overall	9	86.8% (< 0.0001)	1.95	0.708	3.198	0.002

 Table 4
 Subgroup analysis of treatment effect of scaffold embedded chABC on motor function recovery after SCI compared to non-treated animals

CI confidence interval, SCI spinal cord injury, SMD standardized mean difference

articles showed a high risk of bias in term of the reasons to exclude animals from the study (Table 3).

Results of meta-analysis

In the current study, publication bias was observed in 9 studies describing the effects of a scaffold embedded with chABC on the motor function recovery test of animals after SCI (p = 0.02) (Fig. 2).

The results of the meta-analysis showed that embedding chABC into a scaffold had a significant effect on the recovery of motor function (SMD=1.95; 95% CI 0.71– 3.2; p=0.002) (Fig. 3). The results also showed that there was a significant heterogeneity between the studies of motor function recovery (I^2 =86.8%, p<0.001).

In the subgroup analysis (Table 4), it was found that the following factors relevant to the SCI model had no statistical effect on the outcome. These were contusion-compression (SMD=1.4; 95% CI:-0.52 to 3.31; p = 0.15) or transection-hemisection (SMD = 2.458; 95% CI: 0.688–4.228; p=0.006); the follow-up time, which was <8 weeks (SMD = 1.55; 95% CI: 0.572 - 2.527; p=0.002) or ≥ 8 weeks (SMD=3.11; 95% CI: -0.89 to 7.1; p = 0.128); severity of injury which was severe (SMD = 2.694; 95% CI: 0.793-4.595; p = 0.005) or moderate (SMD = 1.146; 95% CI: -0.567 to 2.858; p = 0.19); or the transplantation method which was by injection (SMD = 1.45; 95% CI - 0.093 to 2.997; p = 0.0.065) or by implantation (SMD=2.801; 95% CI 0.399-5.204; p=0.022). None of these factors had any influence on the effects of scaffold embedded chABC on the motor function recovery after SCI (Table 4), and in all the sub groups the scaffold embedded chABC had a significant effect compared to the untreated animals.

Discussion

In this article, the effect of the chABC enzyme embedded in a scaffold on the treatment of SCI was systematically investigated. The results showed that the chABC embedded in a scaffold could significantly improve the functional recovery compared to untreated animals.

Many pre-clinical articles have examined the effects of chABC injection on the improvement of functional recovery after SCI [58–62]. The results of a meta-analysis showed that the enzyme chABC injected alone had a moderate effect on the functional recovery post-SCI (SMD = 0.90; 95% CI 0.61 to 1.20; p < 0.001) [40].

The results of the present study that analyzed all articles that employed chABC embedded in a hydrogel or a scaffold, showed that its effect on the functional recovery post-SCI was stronger and more significant than chABC alone. Based on the result, none of the factors involved in the SCI model, SCI severity, location of the injury, type of transplant, follow-up time, influenced on the therapeutic effect that according to the small number of studies makes it impossible to accept this result with certainty. Treatment of a damaged spinal cord with chABC is designed to attenuate the inhibitory effects of CSPGs that accumulate in the lesion site, and is expected to improve overall functional recovery. Injection of the free enzyme is the most common method to administer chABC, but the disadvantage of the injection at the lesion site, is that the injected chABC is subject to rapid degradation by the host enzyme and body temperature [63, 64], and also spreads away from the site of injury [41]. There are major limitations to the path of clinical treatment through direct injection of chABC; ChABC after 1 h (h) of incubation at 37 °C fails 50% of its enzymatic activity.

Therefore, to circumvent this barrier, several injections of ChABC, infusion through mini-pumps/catheters or use of scaffolds have been used to provide stable continuous local delivery of fresh chABC in vivo [33]. The result of this meta-analysis showed when the chABC is locally delivered by embedding it into a scaffold which allows slow-release, this approach can maintain higher levels of the bioactive enzyme at the lesion site for a longer time and thus increase the beneficial effect on neuronal regeneration and functional recovery [60, 65].

Although our analysis was based on 9 articles, and the number of articles could be seen as one of the limitations, nevertheless the results can pave the way for the use of scaffold-embedded chABC to treat SCI and suggests this route of administration is superior to injection of chABC alone.

Acknowledgements

Not applicable.

Author contributions

Study design: FR. Article screening: ASH, AZ. Data extraction: ASH, AZ, ZB. Data analysis: FR, MY. Writing: FR, SM, MRH. All authors reviewed the manuscript. All authors read and approved the final manuscript.

Funding

FR was supported by the IRAN University of Medical Sciences, Grant No. 98-4-99-16855. MRH was supported by US NIH Grants R01Al050875 and R21Al121700.

Data availability

The findings of this study are derived from data that are available upon request from the responsible author (FR).

Declarations

Ethics approval and consent to participate

Not applicable.

Competing interests

MRH states the possible conflicts of interest in Additional file. The other authors declare no conflict of interest.

Author details

¹ Department of Medical Nanotechnology, Faculty of Advanced Technologies in Medicine, Iran University of Medical Sciences, Tehran, Iran. ²Department of Physiology, School of Medicine, Iran University of Medical Sciences, Tehran, Iran. ³Laser Research Centre, Faculty of Health Science, University of Johannesburg, Doornfontein 2028, South Africa. ⁴Radiation Biology Research Center, Iran University of Medical Sciences, Tehran, Iran. ⁵Physiology Research Center, Iran University of Medical Sciences, Tehran, Iran. ⁶Physiology Research Center, Institute of Neuropharmacology, Kerman University of Medical Sciences, Kerman, Iran.

Received: 28 February 2022 Accepted: 26 July 2022 Published online: 28 October 2022

References

- Ko H. Revisit spinal shock: pattern of reflex evolution during spinal shock. Korean J Neurotrauma. 2018;14:47.
- Liau LL, Looi QH, Chia WC, Subramaniam T, Ng MH, Law JX. Treatment of spinal cord injury with mesenchymal stem cells. Cell Biosci. 2020;10:112.

- Alizadeh A, Dyck SM, Karimi-Abdolrezaee S. Traumatic spinal cord injury: an overview of pathophysiology, models and acute injury mechanisms. Front Neurol. 2019;10:282.
- Zeinali A, Mellat A, Rahimdel A, Alahsab A. A study on demographic pattern, causes and level of lesion in the spinal cord injured patients in Yazd City in 2009–2015. Int J Med Res Health Sci. 2016;5:447–51.
- Yousefifard M, Ramezani F, Vaccaro AR, Hosseini M, Rahimi-Movaghar V. The role of intraspinal administration of self-assembled peptide on locomotion recovery after spinal cord injury: a systematic review and meta-analysis study. Neuromodulation. 2022. https://doi.org/10.1016/j. neurom.2022.01.011.
- Kahuripour M, Behroozi Z, Rahimi B, Hamblin MR, Ramezani F. The potential of curcumin for treating spinal cord injury: a meta-analysis study. Nutr Neurosci. 2022. https://doi.org/10.1080/1028415X.2022.2070703.
- Behroozi Z, Rahimi B, Hamblin MR, Nasirinezhad F, Janzadeh A, Ramezani F. Injection of cerium oxide nanoparticles to treat spinal cord injury in rats. J Neuropathol Exp Neurol. 2021. https://doi.org/10.21203/rs.3.rs-547438/v1.
- Ramezani F, Neshasteh-Riz A, Ghadaksaz A, Fazeli SM, Janzadeh A, Hamblin MR. Mechanistic aspects of photobiomodulation therapy in the nervous system. Lasers Med Sci. 2021. https://doi.org/10.1007/ s10103-021-03277-2.
- Ramezani F, Razmgir M, Tanha K, Nasirinezhad F, Neshastehriz A, Bahrami-Ahmadi A, et al. Photobiomodulation for spinal cord injury: a systematic review and meta-analysis. Physiol Behav. 2019;2020(224): 112977.
- Yousefi S, Hojati V, Nasirinezhad F, Ramezani F, Janzadeh A, Vaezi G. The effect of four weeks of low-level laser radiation (660 nm) on movement recovery and fibroblasts invasion. Arch Neurosci. 2019. https://doi.org/10. 5812/ans.87225.
- Tran AP, Warren PM, Silver J. The biology of regeneration failure and success after spinal cord injury. Physiol Rev. 2018;98:881–917.
- Bradbury EJ, Burnside ER. Moving beyond the glial scar for spinal cord repair. Nat Commun. 2019;10:3879.
- Fan B, Wei Z, Yao X, Shi G, Cheng X, Zhou X, et al. Microenvironment imbalance of spinal cord injury. Cell Transplant. 2018;27:853–66.
- 14. Ohtake Y, Smith G, Li S. Reactive astrocyte scar and axon regeneration: suppressor or facilitator? Neural Regen Res. 2016;11:1050.
- Ghosh S, Hui SP. Axonal regeneration in zebrafish spinal cord. Regeneration. 2018;5:43–60.
- Selzer ME, Dobkin BH. Spinal cord injury: a guide for patients and families (American Academy of Neurology Press Quality of Life Guides). AAN PRESS; 2008.
- Zhou P, Guan J, Xu P, Zhao J, Zhang C, Zhang B, et al. Cell therapeutic strategies for spinal cord injury. Adv Wound Care. 2019;8:585–605.
- Orlandin JR, Ambrósio CE, Lara VM. Glial scar-modulation as therapeutic tool in spinal cord injury in animal models. Acta Cir Bras. 2017;32:168–74.
- Gaudet AD, Fonken LK. Glial cells shape pathology and repair after spinal cord injury. Neurotherapeutics. 2018;15:554–77.
- Jones LL, Sajed D, Tuszynski MH. Axonal regeneration through regions of chondroitin sulfate proteoglycan deposition after spinal cord injury: a balance of permissiveness and inhibition. J Neurosci. 2003;23:9276–88.
- Siebert JR, Conta Steencken A, Osterhout DJ. Chondroitin sulfate proteoglycans in the nervous system: inhibitors to repair. Biomed Res Int. 2014;2014:1–15.
- Wanner IB, Deik A, Torres M, Rosendahl A, Neary JT, Lemmon VP, et al. A new in vitro model of the glial scar inhibits axon growth. Glia. 2008;56:1691–709.
- Cheng C-H, Lin C-T, Lee M, Tsai M, Huang W-H, Huang M-C, et al. Local delivery of high-dose chondroitinase ABC in the sub-acute stage promotes axonal outgrowth and functional recovery after complete spinal cord transection. PLoS ONE. 2015;10: e0138705.
- Yick L, Wu W, So K-F, Yip HK, Shum DK. Chondroitinase ABC promotes axonal regeneration of Clarke's neurons after spinal cord injury. NeuroReport. 2000;11:1063–7.
- Nagappan PG, Chen H, Wang D. Neuroregeneration and plasticity: a review of the physiological mechanisms for achieving functional recovery postinjury. Mil Med Res. 2020;7:30.
- Dyck S, Kataria H, Alizadeh A, Santhosh KT, Lang B, Silver J, et al. Perturbing chondroitin sulfate proteoglycan signaling through LAR and PTPσ

receptors promotes a beneficial inflammatory response following spinal cord injury. J Neuroinflamm. 2018;15:90.

- Matrix E, Injury TB. Extracellular matrix and traumatic brain injury. J Neurosci Res. 2018;96:573–88.
- Hussein RK, Mencio CP, Katagiri Y, Brake AM, Geller HM. Role of chondroitin sulfation following spinal cord injury. Front Cell Neurosci. 2020;14:208.
- Laabs TL, Wang H, Katagiri Y, McCann T, Fawcett JW, Geller HM. Inhibiting glycosaminoglycan chain polymerization decreases the inhibitory activity of astrocyte-derived chondroitin sulfate proteoglycans. J Neurosci. 2007;27:14494–501.
- Pakulska MM, Tator CH, Shoichet MS. Local delivery of chondroitinase ABC with or without stromal cell-derived factor 1α promotes functional repair in the injured rat spinal cord. Biomaterials. 2017;134:13–21.
- Warren PM, Andrews MR, Smith M, Bartus K, Bradbury EJ, Verhaagen J, et al. Secretion of a mammalian chondroitinase ABC aids glial integration at PNS/CNS boundaries. Sci Rep. 2020;10:11262.
- Pearson CS, Mencio CP, Barber AC, Martin KR, Geller HM. Identification of a critical sulfation in chondroitin that inhibits axonal regeneration. Elife. 2018;7:1–21.
- Raspa A, Bolla E, Cuscona C, Gelain F. Feasible stabilization of chondroitinase abc enables reduced astrogliosis in a chronic model of spinal cord injury. CNS Neurosci Ther. 2019;25:86–100.
- Yang T, Dai Y, Chen G, Cui S. Dissecting the dual role of the glial scar and scar-forming astrocytes in spinal cord injury. Front Cell Neurosci. 2020;14:78.
- 35. He Y, Liu X, Chen Z. Glial scar—a promising target for improving outcomes after CNS injury. J Mol Neurosci. 2020;70:340–52.
- Xie C, Shen X, Xu X, Liu H, Li F, Lu S, et al. Astrocytic YAP promotes the formation of glia scars and neural regeneration after spinal cord injury. J Neurosci. 2020;40:2644–62.
- 37. Sami A, Selzer ME, Li S. Advances in the signaling pathways downstream of glial-scar axon growth inhibitors. Front Cell Neurosci. 2020;14:174.
- Warren PM, Dickens SM, Gigout S, Fawcett JW, Kwok JCF. Regulation of CNS plasticity through the extracellular matrix. In: Chao MV, editor. The Oxford handbook of developmental neural plasticity. Oxford: Oxford University Press; 2018. p. 1–40.
- Tom VJ, Kadakia R, Santi L, Houlé JD. Administration of chondroitinase ABC rostral or caudal to a spinal cord injury site promotes anatomical but not functional plasticity. J Neurotrauma. 2009;26:2323–33.
- Yousefifard M, Chondroitinase ABC. Administration in locomotion recovery after spinal cord injury: a systematic review and meta-analysis. Basic Clin Neurosci J. 2021. https://doi.org/10.1016/j.expneurol.2007.07.019.
- Lee H, McKeon RJ, Bellamkonda RV. Sustained delivery of thermostabilized chABC enhances axonal sprouting and functional recovery after spinal cord injury. Proc Natl Acad Sci. 2010;107:3340–5.
- Clemons TD, Fitzgerald M, Dunlop SA, Harvey AR, Iyer KS, Stubbs KA. An improved assay for the spectrophotometric determination of chondroitinase ABC activity. New J Chem. 2013;37:1944.
- Pakulska M. Combined delivery of chondroitinase ABC (ChABC) and stromal cell derived factor 1α (SDF1α) for spinal cord regeneration (Doctoral dissertation). 2016.
- Jahandideh A, Noori H, Rahimi B, Hamblin MR, Behroozi Z, Ramezani M, et al. Alginate scaffolds improve functional recovery after spinal cord injury. Eur J Trauma Emerg Surg. 2021;48:1711–21.
- 45. Fan M, Tan H. Biocompatible conjugation for biodegradable hydrogels as drug and cell scaffolds. Cogent Eng. 2020;7:1736407.
- 46. Wang X, Yang Y, Shi Y, Jia F. Editorial: Smart hydrogels in tissue engineering and regenerative medicine. Front Chem. 2020;8.
- Song R, Murphy M, Li C, Ting K, Soo C, Zheng Z. Current development of biodegradable polymeric materials for biomedical applications. Drug Des Dev Ther. 2018;12:3117–45.
- Azizi M, Farahmandghavi F, Joghataei MT, Zandi M, Imani M, Bakhtiari M, et al. ChABC-loaded PLGA nanoparticles: a comprehensive study on biocompatibility, functional recovery, and axonal regeneration in animal model of spinal cord injury. Int J Pharm. 2020;577: 119037.
- Huang Y-C, Hsu S, Chen M, Hsieh C, Kuo W, Cheng H, et al. Controlled release of chondroitinase ABC in chitosan-based scaffolds and PDLLA microspheres. Carbohydr Polym. 2011;84:788–93.
- 50. Colello RJ, Chow WN, Bigbee JW, Lin C, Dalton D, Brown D, et al. The incorporation of growth factor and chondroitinase ABC into an

electrospun scaffold to promote axon regrowth following spinal cord injury. J Tissue Eng Regen Med. 2016;10:656–68.

- Hassannejad Z, Vahedi F, Zadegan SA, Mokhatab M, Rezvan M, Saadat S, et al. Potential variables affecting the quality of animal studies regarding pathophysiology of traumatic spinal cord injuries. Spinal Cord. 2015;54:579–83.
- Cholas R, Hsu H, Spector M.Collagen Scaffolds Incorporating Select Therapeutic Agents to Facilitate aReparative Response in a Standardized Hemiresection Defect in the Rat SpinalCord. Tissue Eng Part A. 2012;18:2158–72.
- 53. Führmann T, Anandakumaran PN, Payne SL, Pakulska MM, Varga BV, Nagy A, Tator C, Shoichet MS. Combined delivery of chondroitinase ABC and human induced pluripotent stem cell-derived neuroepithelial cells promote tissue repair in an animal model of spinal cord injury. Biomed Mater. 2018;13(2):024103. https://doi.org/10.1088/1748-605X/aa96dc.
- 54. Xia T, Huang B, Ni S, Gao L,Wang J, Wang J, et al. The combination of dbcAMP and ChABC with poly(propylenecarbonate) microfibers promote axonal regenerative sprouting and functionalrecovery after spinal cord hemisection injury. Biomed Pharmacother.2017;86:354–62.
- Zhang Y, Gu Z, Qiu G, Song Y. Combination of chondroitinase ABC, glial cell line–derived neurotrophic factor and Nogo A antibody delayedrelease microspheres promotes the functional recovery of spinal cord injury. J Craniofac Surg. 2013;24:2153–7.
- Pan Q, Guo Y, Kong F. Poly (glycerol sebacate) combined with chondroitinase ABC promotes spinal cord repair in rats. 2017;1–8.
- Ni S, Xia T, Li X, Zhu X, Qi H,Huang S, et al. Sustained delivery of chondroitinase ABC by poly(propylene carbonate)-chitosan micron fibers promotes axon regeneration and functional recovery after spinal cord hemisection. Brain Res. 2015;1624:469–78.
- García-Alías G, Barkhuysen S, Buckle M, Fawcett JW. Chondroitinase ABC treatment opens a window of opportunity for task-specific rehabilitation. Nat Neurosci. 2009;12:1145–51.
- Lin R, Kwok JCF, Crespo D, Fawcett JW. Chondroitinase ABC has a longlasting effect on chondroitin sulphate glycosaminoglycan content in the injured rat brain. J Neurochem. 2007;104(2):400–8.
- Song YH, Agrawal NK, Griffin JM, Schmidt CE. Recent advances in nanotherapeutic strategies for spinal cord injury repair. Adv Drug Deliv Rev. 2019;148:38–59.
- Hanna A, et al. Peripheral nerve grafts and chondroitinase ABC application improves functional recovery after complete spinal cord transection. J Neurol Res. 2013;3:85.
- 62. Bukhari N, Torres L, Robinson JK, Tsirka SE. Axonal regrowth after spinal cord injury via chondroitinase and the tissue plasminogen activator (tPA)/ plasmin system. J Neurosci. 2011;31:14931–43.
- Tester NJ, Plaas AH, Howland DR. Effect of body temperature on chondroitinase ABC's ability to cleave chondroitin sulfate glycosaminoglycans. J Neurosci Res. 2007;85:1110–8.
- Liu Y, Wang J, Shibata S, Chio J, Hettiaratchi MH, Fu T, et al. Human oligodendrogenic neural progenitor cells delivered with chondroitinase ABC facilitate functional repair of chronic spinal cord injury. Stem Cell Rep. 2018;11(6):1433–48.
- Hyatt AJT, Wang D, Kwok JC, Fawcett JW, Martin KR. Controlled release of chondroitinase ABC from fi brin gel reduces the level of inhibitory glycosaminoglycan chains in lesioned spinal cord. J Control Release. 2010;147:24–9.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.